

CORRECTION

Open Access



Correction: Perinatal foodborne titanium dioxide exposure-mediated dysbiosis predisposes mice to develop colitis through life

Caroline Carlé¹, Delphine Boucher^{2†}, Luisa Morelli^{3,4†}, Camille Larue⁵, Ekaterina Ovtchinnikova¹, Louise Battut¹, Kawthar Boumessid¹, Melvin Airaud¹, Muriel Quaranta-Nicaise¹, Jean-Luc Ravanat⁶, Gilles Dietrich¹, Sandrine Menard¹, Gérard Eberl^{7,8}, Nicolas Barnich², Emmanuel Mas^{1,9}, Marie Carriere⁶, Ziad Al Nabhani^{3,4*†} and Frédérick Barreau^{1*†}

Correction: Particle and Fibre Toxicology (2023) 20:45
<https://doi.org/10.1186/s12989-023-00555-5>

Following publication of the original article [1], the authors reported some spelling and bibliograph errors.

Below is a table of corrections which have been implemented in the original article.

The original article [1] has been corrected.

[†]Delphine Boucher and Luisa Morelli have contributed equally to this work.

[†]Ziad Al Nabhani, and Frédérick Barreau have contributed equally to this work.

The original article can be found online at <https://doi.org/10.1186/s12989-023-00555-5>.

*Correspondence:

Ziad Al Nabhani
ziad.alnabhani@unibe.ch
Frédérick Barreau
frederick.barreau@inserm.fr

¹ Institut de Recherche en Santé Digestive (IRSD), INSERM UMR-1220, Purpan Hospital, CS60039, University of Toulouse, INSERM, INRAE, ENVT, UPS, 31024 Toulouse Cedex 03, France

² M2ISH, Université Clermont Auvergne, UMR1071 INSERM, USC INRAE 1382, Clermont-Ferrand, France

³ Department of Visceral Surgery and Medicine, Bern University Hospital, University of Bern, 3010 Bern, Switzerland

⁴ Maurice Müller Laboratories, Department for Biomedical Research, University of Bern, 3008 Bern, Switzerland

⁵ Laboratoire Ecologie Fonctionnelle et Environnement, Université de Toulouse, CNRS, Toulouse, France

⁶ Univ. Grenoble-Alpes, CEA, CNRS, IRIG-SyMMES, CIBEST, Grenoble, France

⁷ Institut Pasteur, Microenvironment and Immunity Unit, 75724 Paris, France

⁸ INSERM U1224, Paris, France

⁹ Gastroenterology, Hepatology, Nutrition, Diabetology and Hereditary Metabolic Diseases Unit, Hôpital des Enfants, CHU de Toulouse, 31300 Toulouse, France



Section	Originally published text	Corrected text
Abstract	Perinatal exposure to titanium dioxide (TiO ₂), as a foodborne particle, may influence the intestinal barrier function and the susceptibility to develop inflammatory bowel disease (IBD) later in life	Perinatal exposure to titanium dioxide (TiO ₂), as a foodborne particle, may influence the intestinal barrier function and the susceptibility to develop inflammatory bowel diseases (IBD) later in life
Background	<p>A significant number of human chronic diseases (inflammatory, metabolic ...) is linked to a deficiency of the IBF and some of them, like IBD, exhibit alterations of the four IBF's compartments [8, 9]</p> <p>To evaluate this hypothesis, we exposed pregnant female C57BL/6 mice to 9 mg E171/kg b.w./day via their drinking water, from the beginning of gestation until 3 weeks postdelivery</p> <p>This exposure concentration is in the lower range of the estimated daily exposure of human adults, which ranges between 5.5 and 10.4 mg/kg b.w./day according to EFSA's estimations [ref 35]</p> <p>When considering the guidances on dose conversion between human and animal exposure, such as the Nair and Jacob practice guide or FDA's guidelines, we previously estimated that doses up to 50–60 mg/kg b.w./day in mice would be realistic [ref notre revue PFT] confirming that the dose used in the present study can be considered as a low exposure dose</p>	<p>significant number of human chronic diseases (inflammatory, metabolic ...) is linked to a deficiency of the IBF and some of them, like IBD, exhibit alterations of the three IBF's compartments [8, 9]</p> <p>To evaluate this hypothesis, we exposed pregnant female C57BL/6 mice to 9 mg E171/kg b.w./day via their drinking water, from the beginning of gestation until 4 weeks postdelivery</p> <p>This exposure concentration is in the lower range of the estimated daily exposure of human adults, which ranges between 5.5 and 10.4 mg/kg b.w./day according to EFSA's estimations [29]</p> <p>When considering the guidances on dose conversion between human and animal exposure, such as the Nair and Jacob practice guide or FDA's guidelines, we previously estimated that doses up to 50–60 mg/kg b.w./day in mice would be realistic [14] confirming that the dose used in the present study can be considered as a low exposure dose</p>
Results	Figure 1 Abilities of foodborne TiO ₂ to translocate across the human barriers. A–G Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day)	Figure 1 Abilities of foodborne TiO ₂ to translocate across the human barriers. A–G Wild type female mice have been exposed to TiO ₂ (9 mg/Kg ofBW/Day)

Section	Originally published text	Corrected text
	Since gut microbiota is described to modulate the intestinal epithelium homeostasis [29, 30], we investigated if perinatal exposure to foodborne TiO ₂	Since gut microbiota is described to modulate the intestinal epithelium homeostasis [30, 31], we investigated if perinatal exposure to foodborne TiO ₂
	In addition, the expression of myosin light chain kinase (<i>Mylk</i>), a master regulator of the tight junction opening [31], was increased by perinatal exposure	In addition, the expression of myosin light chain kinase (<i>Mylk</i>), a master regulator of the tight junction opening [32], was increased by perinatal exposure
	Figure 2 Impact of perinatal exposure to foodborne TiO ₂ on colonic microbiota at days 30. A–E Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day) during the perinatal period including gestational and lactating periods. Then at days 30 after birth, pups have been sacrificed and the structure of the colonic mucosa-associated microbiota has been monitored by 16S rRNA gene sequencing (B–E) C–E Composition of colonic microbiota at phyla level (C) and Fold changes 2 for bacterial genera significantly perturbed (D and E) from exposed or non-exposed mice to foodborne TiO ₂ at day 30 after birth	Figure 2 Impact of perinatal exposure to foodborne TiO ₂ on colonic microbiota at day 30. A–D Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day) during the perinatal period including gestational and lactating periods. Then at day 30 after birth, pups have been sacrificed and the structure of the colonic mucosa-associated microbiota has been monitored by 16S rRNA gene sequencing (B–D) C–D Composition of colonic microbiota at phyla level (C) and Fold changes 2 for bacterial genera significantly perturbed (D) from exposed or non-exposed mice to foodborne TiO ₂ at day 30 after birth
	At days 50 after birth, TiO ₂ exposure only increased the level of <i>Muc2</i> (Additional file 5: Fig.S5A, B)	At days 50 after birth, TiO ₂ exposure only increased the level of <i>Muc2</i> (Additional file 5: Fig. S5A–C)
	At days 50 after birth, TiO ₂ exposure only increased the level of <i>Muc2</i> (Additional file 5: Fig. S5A, E)	At days 50 after birth, TiO ₂ exposure only increased the level of <i>Muc2</i> (Additional file 5: Fig. S5A)
	Since perinatal exposure to TiO ₂ altered the functionality of the colonic epithelium, we then monitored its effects on the intestinal epithelial stem cells (IESC) homeostasis (Fig. 3D–F; Additional file 5: Fig. S3D–F)	Since perinatal exposure to TiO ₂ altered the functionality of the colonic epithelium, we then monitored its effects on the intestinal epithelial stem cells (IESC) homeostasis (Fig. 4D–F; Additional file 5: Fig. S4D–F)

Section	Originally published text	Corrected text
	At day 50, mice exposed to TiO ₂ had an increased mRNA levels of colonic CD44, Leucine-rich repeat-containing G-protein coupled receptor 5 (<i>Lgr5</i>), Achaete-scute complex homolog 2 (<i>Ascl2</i>) and Musashi RNA-binding protein 1 (<i>Musashi</i>), three markers of CBC, Telomerase reverse transcriptase (<i>Tert</i>) and Homeodomain-only protein X (<i>Hopx</i>), two markers of +4 stem cells and the marker of non-canonical wnt pathway (<i>wnt5</i> , involved in inflammatory pathway) (Additional file 3: Fig. S3D) but	At day 50, mice exposed to TiO ₂ had an increased mRNA levels of colonic CD44, Leucine-rich repeat-containing G-protein coupled receptor 5 (<i>Lgr5</i>), Achaete-scute complex homolog 2 (<i>Ascl2</i>) and Musashi RNA-binding protein 1 (<i>Musashi</i>), three markers of CBC, Telomerase reverse transcriptase (<i>Tert</i>) and Homeodomain-only protein X (<i>Hopx</i>), two markers of +4 stem cells and the marker of non-canonical wnt pathway (<i>wnt5</i> , involved in inflammatory pathway) (Additional file 4: Fig. S4D) but
	Figure 3 Impact of perinatal exposure to foodborne TiO ₂ on colonic epithelium at day 30. A–D Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day)	Figure 3 Impact of perinatal exposure to foodborne TiO ₂ on colonic epithelium at day 30. A–D Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day)
	We observed a significant reduction of organoid growth at day 9 post-organoid culture obtained from TiO ₂ -exposed mice compared to control at day 30 (Fig. 3E) but the survival of colonic organoids was similar between both TiO ₂ -treated and untreated group (Fig. 3F)	We observed a significant reduction of organoid growth at day 9 post-organoid culture obtained from TiO ₂ -exposed mice compared to control at day 30 (Fig. 3F) but the survival of colonic organoids was similar between both TiO ₂ -treated and untreated group (Fig. 3E)
	Finally, since oxidative stress and/or DNA methylation are well known to regulate gene expression, we monitored the impact of exposure to TiO ₂ on the oxidative balance as well as DNA methylation of the colonic epithelium (Fig. 3G, H; Additional file 4: Fig. S4H)	Finally, since oxidative stress and/or DNA methylation are well known to regulate gene expression, we monitored the impact of exposure to TiO ₂ on the oxidative balance as well as DNA methylation of the colonic epithelium (Fig. 3G, H; Additional file 4: Fig. S4G)
	In this objective, we used 8-oxo-dGuo as a biomarker of DNA oxidation, this lesion being also considered as a marker of oxidative stress [32] and being quantifiable with a high sensitivity using methods such as HPLC-tandem mass spectrometry [33]	In this objective, we used 8-oxo-dGuo as a biomarker of DNA oxidation, this lesion being also considered as a marker of oxidative stress [33] and being quantifiable with a high sensitivity using methods such as HPLC-tandem mass spectrometry [34]

Section	Originally published text	Corrected text
	As a DNA methylation biomarker, we quantified 5-methyl-2'-deoxycytidine, i.e., 5-Me-dC, as it is the predominant methylation site in mammalian genomes and it shows the highest biological significance as it modulates the binding of transcription factors to DNA [34, 35]	As a DNA methylation biomarker, we quantified 5-methyl-2'-deoxycytidine, i.e., 5-Me-dC, as it is the predominant methylation site in mammalian genomes and it shows the highest biological significance as it modulates the binding of transcription factors to DNA [29, 35]
	Figure 4 Impact of perinatal exposure to TiO ₂ foodborne on intestinal immune system. A–E Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day) during	Figure 4 Impact of perinatal exposure to TiO ₂ foodborne on intestinal immune system. A–D Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day) during
	In contrast to those observed in colon of young mice, perinatal exposure to TiO ₂ did not affect the mRNA level of <i>Il23</i> while it increased the expression of <i>Il1b</i> , <i>Il6</i> , <i>Il10</i> , <i>Il22</i> and <i>Tnfa</i> (Additional file 6: Fig. S6C)	In contrast to those observed in colon of young mice, perinatal exposure to TiO ₂ did not affect the mRNA level of <i>Il23</i> at day 50 while it increased the expression of <i>Il1b</i> , <i>Il6</i> , <i>Il10</i> , <i>Il22</i> , <i>Tnfa</i> and <i>Ifng</i> (Additional file 6: Fig. S6C)
	However, at protein level, perinatal exposure to TiO ₂ increased the colonic cytokines expression of <i>Tnfa</i> , <i>Ifny</i> , <i>IL-12</i> and <i>IL-1β</i> (Fig. 4A)	However, at protein level, perinatal exposure to TiO ₂ increased the colonic cytokines expression of <i>Tnfa</i> , <i>Ifny</i> , <i>IL-12</i> and <i>IL-1β</i> (Fig. 4A) at day 30
	Regarding colonic immune cell populations, flow cytometry experiments on the lamina propria from colon of mice (day 50) evidenced that perinatal exposure to TiO ₂ increased the percentage of myeloid cells (CD11 ⁺),	Regarding colonic immune cell populations, flow cytometry experiments on the lamina propria from colon of mice (day 50) evidenced that perinatal exposure to TiO ₂ increased the percentage of myeloid cells (CD11b ⁺),
	Finally, the reduced percentage of B cells in the lamina propria was associated with reduced faecal levels of IgA, but not IgG at both days 30 and 50 after birth (Fig. 4D; Additional file 5: Fig. S5D)	Finally, the reduced percentage of B cells in the lamina propria was associated with reduced faecal levels of IgA, but not IgG at both days 30 and 50 after birth (Fig. 4B–D; Additional file 5: Fig. S5D)
	Since gut microbiota dysbiosis has been shown to alter the gut homeostasis [7, 29, 38],	Since gut microbiota dysbiosis has been shown to alter the gut homeostasis [7, 30, 38],

Section	Originally published text	Corrected text
	Six weeks after microbiota transfer, permeability and mRNA levels of <i>Occludin</i> , <i>Tjp1</i> , <i>Tjp2</i> and <i>Mylk</i> as well as <i>Il1b</i> , <i>Il12</i> , <i>Tnfa</i> and <i>Ifng</i> were assessed (Fig. 5B, C). As	Six weeks after microbiota transfer, permeability and mRNA levels of <i>Occludin</i> , <i>Tjp1</i> , <i>Tjp2</i> and <i>Mylk</i> as well as <i>Il1b</i> , <i>Il12</i> , <i>Tnfa</i> and <i>Ifng</i> were assessed (Fig. 5B–D). As
	As illustrated in Fig. 5B, the transfer of T iO ₂ -triggered microbiota dysbiosis to healthy germ-free mice led to significantly increased paracellular intestinal permeability (Fig. 5B), increased mRNA level of <i>Mylk</i> , and reduced mRNA level of <i>Tjp1</i> and <i>Tjp2</i> (Fig. 5C)	As illustrated in Fig. 5B, the transfer of T iO ₂ -triggered microbiota dysbiosis to healthy germ-free mice led to significantly increased paracellular intestinal permeability (Fig. 5B), increased mRNA level of <i>Mylk</i> , and reduced mRNA level of <i>Tjp1</i> and <i>Tjp2</i> (Fig. 5C) in offspring at day 30
	We observed that alteration of homeostasis of the colonic mucosa related to early life exposure to TiO ₂ O ₂ did not persist until adult 17 weeks of age as monitored for permeability, cytokine and other inflammatory markers i. e. in the group unchallenged for DSS mice exposed to TiO ₂ superpose with mice unexposed (Fig. 6; Additional file 7: Fig. S7A)	We observed that alteration of homeostasis of the colonic mucosa related to early life exposure to TiO ₂ did not persist until adult 17 weeks of age as monitored for permeability, cytokine and other inflammatory markers i. e. in the group unchallenged for DSS mice exposed to TiO ₂ superpose with mice unexposed (Fig. 6; Additional file 7: Fig. S7)
	However, as illustrated in Fig. 6B–H, perinatal exposure to TiO ₂ enhanced significantly the loss of body weight and the DAI induced by DSS	However, as illustrated in Fig. 6B–G, perinatal exposure to TiO ₂ enhanced significantly the loss of body weight and the DAI induced by DSS. Perinatal
	Figure 6 Impact of perinatal exposure to foodborne TiO ₂ on susceptibility to develop colitis later in life. A–G Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day) during the perinatal period including gestational and lactating periods (A)	Figure 6 Impact of perinatal exposure to foodborne TiO ₂ on susceptibility to develop colitis later in life. A–G Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day) during the perinatal period including gestational and lactating periods (A)
	Perinatal exposure to TiO ₂ also exacerbated the colitis, as evidenced by a reduced colon length associated with increased colonic mRNA expression and protein levels of IL-1β, IL-4, IL-12, IL-13, IFNγ and TNF-α (Additional file 6: Fig. S6A and additional File 7: FigS7E)	Perinatal exposure to TiO ₂ also exacerbated the colitis, as evidenced by a reduced colon length associated with increased colonic mRNA expression and protein levels of IL-1β, IL-4, IL-12, IL-13, IFNγ and TNF-α (Additional file 7: Fig. S7)

Section	Originally published text	Corrected text
	Perinatal exposure to TiO ₂ also aggravated significantly the alterations of intestinal permeability, as evidenced by an increased Dextran-FITC flux, mRNA expression of MLCK and a reduced mRNA level of <i>Tjp1</i> (Fig. 6G)	Perinatal exposure to TiO ₂ also aggravated significantly the alterations of intestinal permeability, as evidenced by an increased 4 kDa Dextran-FITC flux, mRNA expression of MLCK and a reduced mRNA level of <i>Tjp1</i> (Fig. 6G)
	In contrast, at the 17th week of life, there was no longer any significant difference in terms of permeability, cytokine or other inflammatory markers i. e. in the group unchallenged for DSS mice exposed to TiO ₂ superpose with mice unexposed (Fig. 7D–H)	In contrast, at the 17th week of life, there was no longer any significant difference in terms of permeability, cytokine or other inflammatory markers i. e. in the group unchallenged for DSS mice exposed to TiO ₂ superpose with mice unexposed (Fig. 7E–G)
	The colitis was exacerbated in these animals, as evidenced by a reduced colon length associated with increased colonic mRNA expression and protein levels of IL-1β, IL-4, IL-12, IL-13, IFNγ and TNF-α (Additional file 8: Fig. S8 A and Additional file 7: Fig. S7E)	The colitis was exacerbated in these animals, as evidenced by a reduced colon length associated with increased colonic mRNA expression and protein levels of IL-1β, IL-4, IL-12, IL-13, IFNγ and TNF-α (Additional file 8: Fig. S8 and file 7: Fig. 7E)
Discussion	In this study, authors evidenced that foodborne TiO ₂ particles were able to cross the cotyledon of human placenta while no data are available concerning their potential in vivo passage [42] Moreover, the presence of Ti in the meconium do not indicate if its passage underwent during gestation and/or the beginning of suckling	In this study, authors evidenced that foodborne TiO ₂ particles were able to cross the cotyledon of human placenta while no data are available concerning their potential in vivo passage [42]. Moreover, the presence of Ti in the meconium does not indicate if its passage underwent during gestation and/or the beginning of suckling
	This bacteria, which resides in the intestinal mucus layer harbors some virulence traits (type VI secretion system and putative effector proteins) [43], which can trigger CD-like disease in the presence of impaired clearance of the bacterium by innate immunity [44]	This bacteria, which resides in the intestinal mucus layer harbors some virulence traits (type VI secretion system and putative effector proteins) [43], which can trigger IBD-like disease in the presence of impaired clearance of the bacterium by innate immunity [44]

Section	Originally published text	Corrected text
Methods	The deleterious impact of this microbiota dysbiosis is consistent with other microbiota dysbiosis described to affect the intestinal homeostasis then favouring the development of both inflammation and cancer [29, 47, 48]	The deleterious impact of this microbiota dysbiosis is consistent with other microbiota dysbiosis described to affect the intestinal homeostasis then favouring the development of both inflammation and cancer [30, 47, 48]
	these altered mRNA expressions are probably induced and/or linked to the inflammatory context (increased levels of Tnfa, Ifny, IL-12 and IL-1β) of the intestinal epithelium perinatally exposed to TiO ₂	these altered mRNA expressions are probably induced and/or linked to the inflammatory context (increased levels of Tnfa, Ifny, IL-12 and IL-1β) of the intestinal epithelium perinatally exposed to TiO ₂
	Nevertheless, a recent study has reported that microbiota was able to modulate the epigenetic marks on DNA [57]	Nevertheless, a recent study has reported that microbiota was able to modulate the epigenetic marks on DNA [57]
	In more details, 100 days of TiO ₂ exposure slightly increase the dendritic cell frequency while it reduces the regulatory T-cells in Peyer's patches [21]	In more details, 100 days of TiO ₂ exposure slightly increases the dendritic cell frequency while it reduces the regulatory T-cells in Peyer's patches [21]
	Pregnant C57BL/6 wild type female mice were exposed to food additive titanium particles (E171; 9 mg/kg of body weight/day) via drinking water until 3 weeks post-delivery and their offspring was analysed at post-natal day (PND) 30 weaning or maintained under such exposure until PND50	Pregnant C57BL/6 wild type female mice were exposed to food additive titanium particles (E171; 9 mg/kg of body weight/day) via drinking water until 4 weeks post-delivery and their offspring was analysed at post-natal day (PND) 30 weaning or maintained under such exposure until PND50
	Mice were gavaged with FD4 (10 mg/100 μL per mice; Sigma) 4 h before the sacrifice [60]	Mice were gavaged with FD4 (10 mg/100 μL per mice; Sigma) 3 h before the sacrifice [60]
	Permeability was assessed by measuring the mucosal-to-serosal flux of FD4 [30]	Permeability was assessed by measuring the mucosal-to-serosal flux of FD4 [31]
	Organoid stem cell survival (number of organoids formed), and growth capacity (organoid area (μm ²)) were followed three, six, nine and twelve days after plating with a wide field transmission microscope (Apotome Zeiss, 10X lens)	Organoid stem cell survival (number of organoids formed), and growth capacity (organoid area (μm ²)) were followed three, six and nine days after plating with a wide field transmission microscope (Apotome Zeiss, 10X lens)

Section	Originally published text	Corrected text
Supplementary Information	Additional file 1. Fig. S1: Impact of perinatal exposure to foodborne TiO ₂ O ₂ on the composition of chemical element of foetus, spleen and liver from females and pups. (A-C) Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day)	Additional file 1. Fig. S1: Impact of perinatal exposure to foodborne TiO ₂ on the composition of chemical element of foetus, spleen and liver from females and pups. (A-C) Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day)
	(A and B) Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day) during the perinatal period including gestational and lactating periods	(A and B) Wild type female mice have been exposed to TiO ₂ O ₂ (9 mg/Kg of BW/Day) during the perinatal period including gestational and lactating periods
	A-E Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day) during the perinatal period including gestational and lactating periods	A-E Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day) during the perinatal period including gestational and lactating periods
	Weaning pups were also exposed to TiO ₂ (9 mg/BW/Day) until day 50 after birth (A)	Weaning pups were also exposed to TiO ₂ (9 mg/Kg of BW/Day) until day 50 after birth (A)
	Then at day 50 after birth, pups have been sacrificed and the structure of the colonic mucosa-associated microbiota has been monitored by 16S rRNA gene sequencing (B-E)	Then at day 50 after birth, pups have been sacrificed and the structure of the colonic mucosa-associated microbiota has been monitored by 16S rRNA gene sequencing (B-D)
	(C-E) Composition of colonic microbiota at phyla level (C) and Fold changes 2 for bacterial genera significantly perturbed (D and E) from exposed or non-exposed mice to foodborne TiV at day and 50 after birth	C-D) Composition of colonic microbiota at phyla level (C) and Fold changes 2 for bacterial genera significantly perturbed (D) from exposed or non-exposed mice to foodborne TiO ₂ at day and 50 after birth
	Additional file 4. Fig. S4: Impact of perinatal exposure to foodborne TiO ₂ on colonic epithelium at day 50. (A-D) Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day) during the perinatal period including gestational and lactating periods. Weaning pups were also exposed to TiO ₂ (9 mg/BW/Day) until day 50 after birth (A-D)	Additional file 4. Fig. S4: Impact of perinatal exposure to foodborne TiO ₂ on colonic epithelium at day 50. (A-D) Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day) during the perinatal period including gestational and lactating periods. Weaning pups were also exposed to TiO ₂ (9 mg/Kg of BW/Day) until day 50 after birth (A-D)

Section	Originally published text	Corrected text
	(A-D) Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day) during the perinatal period including gestational and lactating periods. Then, at days 30 or 50 after birth, pups have been sacrificed and several parameters including colonic mRNA expression of mucin 2 (<i>Muc2</i>), mucin 3 (<i>Muc3</i>), mucin 4 (<i>Muc4</i>) and Trefoil factor 3 (<i>Tff3</i>) (A), faecal levels of lysozyme (B) and IgG (C)	(A-D) Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day) during the perinatal period including gestational and lactating periods. Then, at days 30 or 50 after birth, pups have been sacrificed and several parameters including colonic mRNA expression of mucin 2 (<i>Muc2</i>), mucin 3 (<i>Muc3</i>), mucin 4 (<i>Muc4</i>) and Trefoil factor 3 (<i>Tff3</i>) (A, B), faecal levels of lysozyme (C) and IgG (D)
	(A-C) Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day) during the perinatal period including gestational and lactating periods. Weaning pups were also exposed to TiO ₂ (9 mg/BW/Day) until day 50 after birth	(A-C) Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day) during the perinatal period including gestational and lactating periods. Weaning pups were also exposed to TiO ₂ (9 mg/Kg of BW/Day) until day 50 after birth
	Wild type female mice have been exposed to TiO ₂ (9 mg/BW/Day) during the perinatal period including gestational and lactating periods. A	Wild type female mice have been exposed to TiO ₂ (9 mg/Kg of BW/Day) during the perinatal period including gestational and lactating periods. A

Published online: 06 March 2024

Reference

1. Carlé C, Boucher D, Morelli L, et al. Perinatal foodborne titanium dioxide exposure-mediated dysbiosis predisposes mice to develop colitis through life. *Part Fibre Toxicol.* 2023;20:45. <https://doi.org/10.1186/s12989-023-00555-5>.

Publisher’s Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.